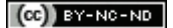


# D-dimer: A Marker of Severity in COVID-19

B SARAVANAN<sup>1</sup>, S VASUKI<sup>2</sup>, BM PABITHADEVI<sup>3</sup>, M SARADHA<sup>4</sup>, R RASKIN ERUSAN<sup>5</sup>,  
S ALAGESAN<sup>6</sup>, SHANTARAMAN KALYANARAMAN<sup>7</sup>



## ABSTRACT

**Introduction:** The ever-growing number of COVID-19 patients stresses upon the need to identify effective yet readily available predictors of disease severity to ensure better clinical outcomes. D-dimer is a fibrin specific degradation product derived by enzymatic action of plasmin on factor XIIIa cross-linked fibrin. It serves as an ideal marker for activation of coagulation and fibrinolytic pathways. Identification of coagulopathy as an important complication in COVID-19 patients has brought to focus D-dimer as a possible predictor of clinical severity in patients.

**Aim:** In this study, we analysed the role of D-dimer levels in assessing the clinical severity of the COVID-19 patients.

**Materials and Methods:** We enrolled 217 in-patients of Tirunelveli Medical College in this single centre observational study and classified them into asymptomatic, mild, moderate and severe according to “Clinical Management Protocol: COVID 19”, by the Ministry of Health and Family Welfare and Director General of Health Services. D-dimer was estimated in the separated plasma, using latex based assay using semi automated coagulation analyser. Data were presented as

percentages for categorical variables and median±Inter Quartile Range (IQR) for continuous variables. Chi-square test was used to compare the D-dimer values between symptomatic and asymptomatic groups. A value of  $p < 0.05$  was considered statistically significant.

**Results:** Among the 217 cases 88.9% were asymptomatic cases, 8.8% presented with mild clinical severity and 2.3% had moderate clinical presentation. In our study population, the Mean±SD and Median±IQR of D-dimer values (in ng/mL) were  $223.4 \pm 230.6$  and  $157.0 \pm 187.7$ , respectively. The mean D-dimer value was found to increase as the category of our study group ascended from asymptomatic patients to mild and moderate clinical cases. It was noted that 91.1% of the cases who had D-dimer values  $< 500$  ng/mL were asymptomatic. Also, the odds of patients with high levels of D-dimer being clinically symptomatic was 5.5 times more than the odds of patients with D-dimer levels  $< 500$  ng/mL.

**Conclusion:** Elevation of D-dimer levels associated with the severity of clinical course of patients infected with SARS CoV-2 when compared to patients with mild or asymptomatic clinical presentations.

**Keywords:** Endothelial dysfunction, Factor XIIIa, Fibrinogen, Severity

## INTRODUCTION

As of 25<sup>th</sup> September 2020, the ongoing pandemic of Coronavirus Disease 2019 (COVID-19) has affected 32,110,656 people worldwide including 980,031 deaths. In India alone, around 5,818,570 have contracted the disease till that date with mortality reaching 92,290 [1]. The Indian Ministry of Health and Family Welfare and Director General of Health Services published “Clinical Management protocol: COVID 19” on 27<sup>th</sup> June 2020, which classifies the disease into mild, moderate and severe cases based on clinical manifestations [2]. Earlier reports from China suggest that approximately 26.1-32.0% of confirmed cases could progress to severe or critical cases [3,4]. The mortality rate is higher among patients with severe and critical illness than those with mild to moderate disease. The ever-growing number of patients stresses upon the need to identify effective yet readily available predictors of disease severity to ensure better clinical outcomes. Researchers have identified coagulopathy as an important complication in COVID-19 patients [5,6]. With this knowledge, researchers are focusing on D-dimer levels in COVID-19 patients with varying degrees of clinical presentation to assess its role as an early predictor of clinical prognosis [6-8].

During haemostasis, coagulation cascade gets activated, resulting in formation of thrombin, a key serine protein involved in clotting [9]. Next thrombin acts upon a 340 kDa soluble glycoprotein called fibrinogen. Fibrinogen molecule is a dimer composed of three pairs of three polypeptide chains which are held together by disulfide bonds. These chains are intertwined in such a way that the fibrinogen molecules consist of a central E domain linked by

coiled-coil regions to two peripheral D domains [10]. The enzymatic activity of thrombin leads to the cleavage of two small fragments, fibrinopeptides A and B, from fibrinogen. This results in formation of fibrin monomer molecules. Also, there is conversion of negative charge present on the E-domain of fibrinogen into positive charge, causing these monomers to spontaneously polymerise into fibrin networks stabilised by hydrogen bonds. Meanwhile thrombin activates Factor XIII, which introduces covalent cross-links between the outer D domains of nearby monomers and the central E-domain of a third one causing further stabilisation of these polymers resulting in insoluble fibrin clots [11,12].

Under physiological conditions, the fibrin thrombi formed during coagulation undergo degradation as soon as they are formed by the fibrinolytic system. This is essential in order to maintain homeostatic balance [13]. The disintegration of fibrin clot begins with the formation of an important fibrinolytic component called plasmin. This is achieved by conversion of fibrin bound plasminogen to plasmin by tissue plasminogen activator, a serine protease released in response to tissue injury [14]. Plasmin mediated proteolysis of fibrinogen and fibrin produces multiple degradation products which are peptide fragments with a wide array of molecular weights [15]. Those fragments derived from fibrin polymers that underwent factor XIIIa mediated crosslinking will have intact covalent bonds bridging adjacent D domains and are called D-dimers [11,16]. Since D-dimer can only be produced when there is formation and degradation of cross-linked fibrin, it serves as a reactive marker of haemostatic balance [17]. D-dimer has a plasma half-life of about 8 hours before clearance by kidneys and reticulo-endothelial system [18]. Normal

reference range of serum D-dimer is 220-740 ng/mL [19]. At present, D-dimer assays are extensively used in management of Deep vein thrombosis, pulmonary embolism, Disseminated Intravascular Coagulation (DIC) and other thromboembolic conditions [11].

Even before the onset of this pandemic, D-dimer was speculated as a marker of organ dysfunction, need for intensive care unit admission and mortality in patients with suspected infection and sepsis [20,21]. Also, increased levels of D-dimer was noted in influenza like illness due to activation of coagulation by certain respiratory viruses [22]. Zhou F et al., and Tang N et al., studies showed D-dimer level >1000 ng/mL results in higher mortality in COVID-19 patients [6,7]. IFCC recommendations published in April, 2020 advocates testing of D-dimer levels in COVID-19 patients [23]. In this study, we analysed the role of D-dimer in assessing the clinical severity of the COVID-19 patients.

## MATERIALS AND METHODS

This is a single centre, cross-sectional study conducted at Government Tirunelveli Medical College, a tertiary care centre in Southern India from March to July 2020. All those who had the contact history with COVID positive cases or clinical signs or symptoms were traced by the Public Health Care Workers and nasopharyngeal swab test was done in the ICMR approved Microbiology Laboratory of Tirunelveli Medical College. Those who tested positive for RT-PCR were admitted in the hospital and **included** in this study irrespective of their age. All those who were negative for RT-PCR were **excluded**. Thus, we enrolled 217 patients (N). This study was approved by the ethics committee of the Institute (REF NO: 1792/PATH/2020) and data were collected retrospectively. Based on the clinical data they were classified, according to the "Clinical Management Protocol: COVID 19" by the Ministry of Health and Family Welfare and Director General of Health Services [2]. The clinical classification was as follows: 1) Asymptomatic: Contacts or patients with travel history tested swab positive by PCR, without any clinical signs like fever, sore throat, dry cough, dyspnoea; 2) Mild: Patients with uncomplicated upper respiratory tract infection may have mild symptoms such as fever, cough, sore throat, nasal congestion, malaise, headache without evidence of breathlessness or Hypoxia (normal saturation); 3) Moderate: Pneumonia with no signs of severe disease. For adolescents or adults with presence of clinical features of dyspnoea and or hypoxia, fever, cough, including SpO<sub>2</sub> <94% (range 90-94%) on room air, Respiratory Rate is more or equal to 24 per minute. For children with presence of clinical features of dyspnoea and or hypoxia, fever, cough, including SpO<sub>2</sub> <94% (range 90-94%) on room air, Respiratory Rate is more or equal to 40 per minute. Fast breathing is defined as (in breaths/min): <2 months: ≥60; 2-11 months: ≥50; 1-5 years: ≥40; 4) Severe: Severe Pneumonia with clinical signs of Pneumonia plus one of the following; respiratory rate >30 breaths/min, severe respiratory distress, SpO<sub>2</sub> <90% on room air for Adolescent or adult and Child with cough or difficulty in breathing, plus at least one of the following: central cyanosis or SpO<sub>2</sub> <90%; severe respiratory distress (e.g., grunting, chest in-drawing); signs of pneumonia with any of the following danger signs: inability to breastfeed or drink, lethargy or unconsciousness, or convulsions; with any other following signs of pneumonia: chest in drawing, fast breathing (in breaths/min): <2 months ≥60; 2-11 months ≥50; 1-5 years ≥40. acute respiratory distress syndrome, sepsis, septic shock and death [2].

## Data Collection and D-dimer Measurement

The patients travel history, exposure history, contact history, demographic data, signs and symptoms, laboratory and Radiological data were collected. Peripheral Venous blood samples were collected as per standard protocol in sodium citrate containers. D dimer was estimated in the separated plasma, using Latex based assay using Semi automated Coagulation analyser (ECL-50, ERBA). Based on the Guan W et al., study (February 2020), the cut-off value for

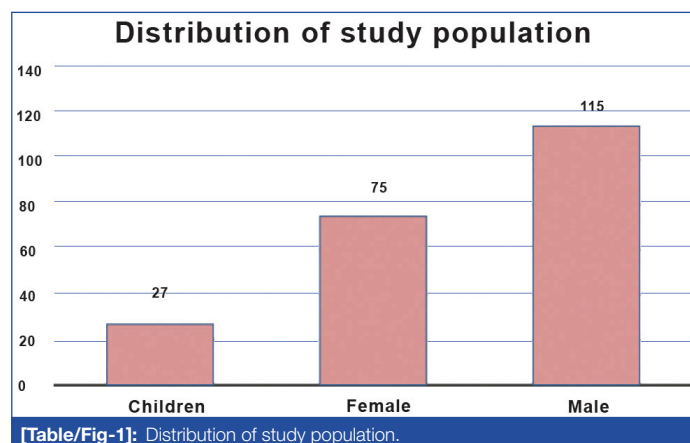
D-dimer level was fixed as 500 ng/mL and the study population are divided into two groups to compare their clinical severity [24]. The throat samples were tested for SARS COV2, with Labgun RT PCR kit (Labgenomics, Republic of Korea) in the hospital laboratory. All collected samples were handled following strict biosafety measures and biomedical waste management guidelines.

## STATISTICAL ANALYSIS

SPSS version 16.0 (SPSS Inc. Chicago, IL) was used. Percentages were used for categorical variables and median±IQR for continuous variables. Chi-square test was used to compare the D-dimer values between symptomatic and asymptomatic groups. A value of p<0.05 was considered statistically significant.

## RESULTS

A total of 217 COVID-19 cases were included in the study population. A 87.5% (n=190) of the study population were adults and among them 52.9% (115) were male and 34.5% (75) were female [Table/Fig-1]. About 27 were children and in 31-40 and 41-50 age group there were 44 patients [Table/Fig-2]. Among the 217 cases 88.9% were asymptomatic cases, 8.8% were presented with mild clinical severity and 2.3% were with moderate clinical presentation [Table/Fig-3]. There was a statistically significant difference in age, Neutrophil count (%), RBC count and NL Ratio between the symptomatic and asymptomatic cases [Table/Fig-4].



[Table/Fig-1]: Distribution of study population.

	Frequency		
	Male	Female	Total
>70 years	2	1	3
61-70 years	14	13	27
51-60 years	21	15	36
41-50 years	26	18	44
31-40 years	26	18	44
21-30 years	17	6	23
13-20 years	9	4	13
<13 years			27

[Table/Fig-2]: Age distribution.

Clinical grading	Frequency	Percent
Asymptomatic cases	193	88.9
Mild clinical presentation	19	8.8
Moderate clinical presentation	5	2.3

[Table/Fig-3]: Distribution of study subjects based on clinical presentation.

In our study population, the Mean±SD and Median±IQR of D-dimer values (in ng/mL) were 223.4±230.6 and 157.0±187.7, respectively. The mean D-dimer value was 212.7 ng/mL in asymptomatic cases which increases to 243.7 ng/mL in mild cases and 525.8 ng/mL in patients with moderate clinical presentation. The mean D-dimer value was found to increase as the category of our study group ascended

	n	Total cases (n=217)		Asymptomatic cases (n=193)		Symptomatic cases (n=24)		Mann-Whitney U test p-value
		Median	IQR*	Median	IQR	Median	IQR	
Age (years)	217	36	27	36	29	42	22	0.04
Blood glucose (mg/dL)	206	88	38.5	88	39	91.5	72.8	0.95
Serum total protein (g/dL)	198	7	0.5	7	0.5	6.9	0.8	0.23
Sodium (mEq/L)	193	140	4	139	4	139.5	3.8	0.80
Potassium (mEq/L)	193	4	0.4	4	0.4	4	0.4	0.45
CK† (U/L)	195	74	90.3	77.6	78	49	64.1	0.09
LDH‡ (U/L)	195	403	146.8	399	153	456	131.3	0.76
Ferritin (ng/mL)	165	112	144.3	107	135	140.5	301.3	0.22
Total WBC§ count (cells/mm³)	206	7550	3400	7500	3100	6900	4700	0.41
Neutrophil (%)	206	56	15	54	15	60.5	21	0.02
Lymphocytes (%)	206	36	15	38	14	38.5	20	0.06
Eosinophil (%)	205	7	5	8	5	6	5	0.07
NL Ratio¶	206	1.5	1.1	1.48	1.03	1.8	2.43	0.03
RBC** (million/mm³)	206	4.8	0.59	4.78	0.59	4.54	0.66	0.01
HB†† (g/dL)	206	13.3	2.6	13.3	2.5	13.3	3.9	0.70
PCV‡‡ (%)	206	38.2	6.6	38.2	6.6	37.8	8.3	0.44
Platelet (Lacs)	203	2.8	1.09	2.82	1.09	2.73	0.99	0.35

**[Table/Fig-4]:** Biochemical and Haematological values in asymptomatic and symptomatic COVID-19 patients.

\*IQR: Inter quartile range; †CK: Creatine kinase; ‡LDH: Lactate dehydrogenase; §WBC: White blood cell; ¶NLRatio: Neutrophil lymphocyte ratio; \*\*RBC: Red blood cell; ††HB: Haemoglobin; ‡‡PCV: Packed cell volume

from asymptomatic patients to mild, moderate clinical cases [Table/Fig-5]. Around 91.1% of the cases who had normal D-dimer values (<500 ng/mL) were asymptomatic cases [Table/Fig-6]. The Chi-square test showed that it was statistically significant and the odds of patients with High level of D-dimer were supposed to be clinically symptomatic cases which were 5.5 times more than that of the odds of patients with normal levels of D-dimer [Table/Fig-7].

Clinical severity	N	Mean (ng/mL)	SD	Median (ng/mL)	IQR
Asymptomatic cases	193	212.7	216.3	151.0	183.9
Mild clinical presentation	19	243.7	212.6	164.0	287.4
Moderate clinical presentation	5	557.8	525.8	556.0	858.4

**[Table/Fig-5]:** D-dimer in Asymptomatic cases, Mild and Moderate cases.

D-dimer category		Patients	
		Symptomatic cases	Asymptomatic cases
Normal (<500 ng/mL)	Count	18	182
	% within D-dimer category	9.0%	91.0%
High (>500 ng/mL)	Count	6	11
	% within D-dimer category	35.3%	64.7%

**[Table/Fig-6]:** Cross tabulation between Clinical severity and D-dimer grading.

	Value	p-value
Pearson chi-square	11.013	0.001
Odds ratio for D-dimer category (High/Normal)	5.5	

**[Table/Fig-7]:** Chi-square Test.

## DISCUSSION

A recent article by Iba T et al., presents a better overview about the possible pathogenesis. Persistent inflammation results in the formation of Interleukin-1 $\beta$  and Interleukin-6 which are known to cause thrombocytosis and hyperfibrinogenemia. In the early stage of disease, inflammation and coagulation are limited to the lungs. But as disease progresses, these features become systemic and later on present as DIC. Alveolar macrophages release urokinase type Plasminogen Activator (u-PA) which promotes local fibrinolysis and D-dimer elevation. Moreover, direct infection of endothelium by the SARS-CoV2 virus via its receptor, Angiotensin-Converting

Enzyme 2 (ACE-2), results in a surge in plasminogen activator release. As disease severity progresses increased fibrinogen levels and activated platelets aggravates procoagulant state. Pulmonary microthrombi formation is promoted by Plasminogen Activator Inhibitor 1 (PAI-1) which leads to suppression of fibrinolysis. In normal vascular endothelium ACE-2 mediates anticoagulant activities. But once SARS-CoV2 virus binds to it, ACE-2 causes cell damage followed by increased expression of tissue factor and downregulation of the protein C system. Ultimately coagulation and thrombotic events may occur even in the absence of secondary complications like tissue hypoxia and superadded infections [25].

Studies conducted in Netherland and France show increased incidence of thrombotic complications in patients with severe disease [26,27]. A retrospective study on 183 patients by Tang N et al., established that 71.4% of nonsurvivors and around 0.6% of the survivors showed features of overt DIC. The median time from admission to development of DIC was 4 days [7]. Autopsy findings published by Wichmann D et al., brings to notice a high incidence of thromboembolism in COVID-19 patients and the need to consider pulmonary embolism in cases of haemodynamic instability [28]. A state of dynamic hypercoagulation as evidenced by the presence of microthrombi in various organs accompanied by reduced platelet levels is seen in COVID patients, especially those with comorbidities. Plasmin mediated hyperactive fibrinolysis may also lead to haemorrhage and marked elevation of circulating fibrin degradation products in such patients [29].

In our study, we noted that the mean D-dimer value increases as the category of our study group moves up from asymptomatic patients to mild, moderate and severe clinical cases. The mean D-dimer value was 212.7 ng/mL in asymptomatic cases which increases to 243.7 ng/mL in mild cases and 525.8 ng/mL in patients with moderate clinical presentation. These findings are in concordance with a multicentre retrospective study conducted in China by Guan W et al., [24]. According to it, 260 of the 560 enrolled patients amounting to 46.4% had elevated D-dimer levels (0.5 mg/L) and this rise in value was more pronounced among severe patients when compared to non-severe ones. They defined the degree of severity of COVID-19 as severe cases if patients admitted with any one of the following major criteria: septic shock with need for vasopressors; respiratory failure requiring mechanical ventilation or any three or more of the following minor



criteria: respiratory rate >30 breaths/min; PaO<sub>2</sub>/FIO<sub>2</sub> ratio <250; multilobar infiltrates; confusion/disorientation; uremia (blood urea nitrogen level >20 mg/dL); leukopenia (white blood cell count, 4,000 cells/mL); thrombocytopenia (platelet count, 100,000/mL); hypothermia (core temperature, 36°C); Hypotension requiring aggressive fluid resuscitation [24]. Study by Tang N et al., in march 2020, showed that patients with severe disease had a 3.5 fold increase in D-dimer levels when compared to those without it along with higher mortality rates [7]. Similar findings were also reported by Yao Y et al., who observed a high level of serum D-dimer level in nonsurvivors when compared to the survivors [30].

A recent guidance report by International Society of Thrombosis and Haemostasis (ISTH) for recognition and management of coagulopathy in COVID-19 states markedly raised D-dimers levels, that is, three-four folds increase as a criteria for admission even though patient has no other severity symptoms [31]. We further established two groups in our study, one whose D-dimer value was above 500 ng/mL and another with values below 500 ng/mL. In our study, population, it was noted that 91.1% of the cases who had normal D-dimer values were asymptomatic. The chi-square test showed that it was statistically significant and the odds of a patient with high D-dimer levels turning out to be clinically symptomatic were 5.5 times more than that of the odds of a patient with normal D-dimer levels.

High D-dimer levels have been reported as one of the risk factors to predict occurrence of acute respiratory distress syndrome and also its progression to death in COVID-19 patients [32]. Wang D et al., noted that patients in need of admission to intensive care units had significantly higher D-dimer levels than those not in need of it [33]. Also, D-dimer levels show a sequential rise with time in nonsurvivors as against values in those who survived [6,33]. Zhang L et al., established a cut-off value of 2.0 µg/mL as an independent predictor for in-hospital death in patients [5].

Initiation of DIC in patients with sepsis is well documented [34]. The pathophysiology behind this is considered to be endothelial dysfunction along with altered immune regulation [35]. Giannis et al., attributed this to endothelial dysfunction characterised by high levels of von Willebrand factor, a procoagulatory state due to activation of tissue factor pathway and toll-like receptor activation causing systemic inflammation in cases of coronavirus infections [36]. Increase in D-dimer levels in COVID-19 may be explained by excess production of thrombin and early shutdown of fibrinolytic pathway secondary to any infectious stimuli as noted in past studies [37]. Another possible explanation for this is hypoxia in severe cases of COVID-19 leading to thrombus formation via increased blood viscosity and hypoxia inducible transcription factor dependent signalling pathway [38]. Some suggest that high levels of circulating proinflammatory cytokines implicated in the cytokine storm noticed in severe cases of COVID-19 might lead to direct activation of coagulation cascade [29].

### Limitation(s)

COVID-19 is a pandemic disease for which prevalence rate is not known. Due to the emerging need for the new classifying factors and new prognostic factors, data over the period of 5 months have been documented in this study. The current study was conducted at a single centre with limited sample size. Hence further study may be needed to extrapolate the findings to wider patient population.

### CONCLUSION(S)

Elevated D-dimer levels associated with the severity of clinical course of patients infected with SARS CoV-2 when compared to patients with mild or asymptomatic clinical presentations. Hence, this reinforces the fact that D-dimer levels at the time of diagnosis helps

to predict the clinical course of the disease and is a definite guide for early management of complications. The prevalence of COVID-19 is not clearly documented and our present knowledge confines to only patients with moderate and severe clinical presentation who reported to hospital for treatment and diagnosed as COVID positive. This enhanced level of D-dimer is a definite indicator of the pathogenetic mechanisms underlying severe clinical presentations and a subject matter of deeper analysis. The role of vW Factor in the increase of D-dimer levels in severely affected COVID-19 patients needs further study.

### Acknowledgement

The investigators acknowledge the support of the Multidisciplinary Research Unit funded by the Department of Health Research, Govt of India. We thank our postgraduate residents for their immense work on data and referencing. We also thank the lab technologists for supporting this study.

### REFERENCES

- [1] WHO Coronavirus Disease (COVID-19) Dashboard [Internet]. [cited 2020 Jul 4]. Available from: <https://covid19.who.int>.
- [2] Clinical Management Protocol for COVID19 dated 27062020.pdf [Internet]. [cited 2020 Jul 18]. Available from: <https://www.mohfw.gov.in/pdf/ClinicalManagementProtocolforCOVID19dated27062020.pdf>.
- [3] Chen N, Zhou M, Dong X, Qu J, Gong F, Han Y, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: A descriptive study. *The Lancet*. 2020;395(10223):507-13.
- [4] Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020;395:497-506.
- [5] Zhang L, Yan X, Fan Q, Liu H, Liu X, Liu Z, et al. D-dimer levels on admission to predict in-hospital mortality in patients with COVID-19. *J Thromb Haemost JTH*. 2020;18(6):1324-29.
- [6] Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: A retrospective cohort study. *The Lancet*. 2020;395(10229):1054-62.
- [7] Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. *J Thromb Haemost JTH*. 2020;18(4):844-47.
- [8] Gao Y, Li T, Han M, Li X, Wu D, Xu Y, et al. Diagnostic utility of clinical laboratory data determinations for patients with the severe COVID-19. *J Med Virol*. 2020;92(7):791-96.
- [9] Ramachandran R, Hansen KK, Hollenberg MD. Proteinase-Activated Receptors. In: Lennarz WJ, Lane MD, editors. *Encyclopedia of Biological Chemistry (Second Edition)* [Internet]. Waltham: Academic Press; 2013. Pp. 60-06. Available from: <http://www.sciencedirect.com/science/article/pii/B9780123786302003984>.
- [10] Hoeprich PDJ, Doolittle RF. Dimeric half-molecules of human fibrinogen are joined through disulfide bonds in an antiparallel orientation. *Biochemistry*. 1983;22(9):2049-55.
- [11] Riley RS, Gilbert AR, Dalton JB, Pai S, McPherson RA. Widely used types and clinical applications of D-Dimer assay. *Lab Med*. 2016;47(2):90-102.
- [12] Wakai A, Gleeson A, Winter D. Role of fibrin D-dimer testing in emergency medicine. *Emerg Med J*. 2003;20(4):319-25.
- [13] Gaffney PJ, Edgell TA, Whittom CM. The haemostatic balance- astrup revisited. *Pathophysiol Haemost Thromb*. 1999;29(1):58-71.
- [14] Longstaff C, Kolev K. Basic mechanisms and regulation of fibrinolysis. *J Thromb Haemost*. 2015;13(S1):S98-105.
- [15] Gaffney PJ. Fibrin degradation products. A review of structures found in vitro and in vivo. *Ann N Y Acad Sci*. 2001;936:594-610.
- [16] Adam SS, Key NS, Greenberg CS. D-dimer antigen: Current concepts and future prospects. *Blood*. 2009;113(13):2878-87.
- [17] Weitz JI, Fredenburgh JC, Eikelboom JW. A Test in Context: D-Dimer. *J Am Coll Cardiol*. 2017;70(19):2411-20.
- [18] Hager K, Platt D. Fibrin degeneration product concentrations (D-Dimers) in the course of ageing. *Gerontology*. 1995;41(3):159-65.
- [19] Kratz A, Pesce MA, Basner RC, Einstein AJ. Appendix: Laboratory Values of Clinical Importance. In: *Harrison's Principles of Internal Medicine*. 19<sup>th</sup> edition. New York: McGraw-Hill Education; 2015. Pp. 2754.
- [20] Garcia-Olivé I, Sintés H, Radua J, Abad Capa J, Rosell A. D-dimer in patients infected with COVID-19 and suspected pulmonary embolism. *Respir Med*. 2020;169:106023.
- [21] Goebel PJ, Williams JB, Gerhardt RT. A pilot study of the performance characteristics of the D-dimer in presumed sepsis. *West J Emerg Med*. 2010;11(2):173-79.
- [22] Wissen MV, Keller TT, Gorp ECMV, Gerdes VEA, Meijers JCM, Doornum GJJV, et al. Acute respiratory tract infection leads to procoagulant changes in human subjects. *J Thromb Haemost*. 2011;9(7):1432-34.
- [23] IFCC Information Guide on COVID-19 Tuesday 14 April updates- IFCC [Internet]. [cited 2020 Jul 4]. Available from: <https://www.ifcc.org/ifcc-news/2020-03-26-ifcc-information-guide-on-covid-19/>.
- [24] Guan W, Ni Z, Hu Y, Liang W, Ou C, He J, et al. Clinical Characteristics of Coronavirus Disease 2019 in China. *N Engl J Med*. 2020;382(18):1708-20.

- [25] Iba T, Levy JH, Connors JM, Warkentin TE, Thachil J, Levi M. The unique characteristics of COVID-19 coagulopathy. *Crit Care*. 2020;24(1):360.
- [26] Julien P, Julien G, Morgan C, Erika P, Thibault D, Fanny L, et al. Pulmonary Embolism in Patients With COVID-19. *Circulation*. 2020;142(2):184-86.
- [27] Klok FA, Kruij MJHA, van der Meer NJM, Arbous MS, Gommers DAMPJ, Kant KM, et al. Incidence of thrombotic complications in critically ill ICU patients with COVID-19. *Thromb Res*. 2020;191:145-47.
- [28] Wichmann D, Sperhake JP, Lütgehetmann M, Steurer S, Edler C, Heinemann A, et al. Autopsy findings and venous thromboembolism in patients with COVID-19. *Ann Intern Med* [Internet]. 2020 May 6 [cited 2020 Jul 18]; Available from: <https://doi.org/10.7326/M20-2003>.
- [29] Ji HL, Zhao R, Matalon S, Matthay MA. Elevated plasmin (ogen) as a common risk factor for COVID-19 susceptibility. *Physiol Rev*. 2020;100(3):1065-75.
- [30] Yao Y, Cao J, Wang Q, Shi Q, Liu K, Luo Z, et al. D-dimer as a biomarker for disease severity and mortality in COVID-19 patients: A case control study. *J Intensive Care*. 2020;8(1):49.
- [31] Thachil J, Tang N, Gando S, Falanga A, Cattaneo M, Levi M, et al. ISTH interim guidance on recognition and management of coagulopathy in COVID-19. *J Thromb Haemost*. 2020;18(5):1023-26.
- [32] Wu C, Chen X, Cai Y, Xia J, Zhou X, Xu S, et al. Risk factors associated with acute respiratory distress syndrome and death in patients with coronavirus disease 2019 Pneumonia in Wuhan, China. *JAMA Intern Med*. 2020;180(7):934-43.
- [33] Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. *JAMA*. 2020;323(11):1061-69.
- [34] Voves C, Wuillemin WA, Zeerleder S. International Society on Thrombosis and Haemostasis score for overt disseminated intravascular coagulation predicts organ dysfunction and fatality in sepsis patients. *Blood Coagul Fibrinolysis Int J Haemost Thromb*. 2006;17(6):445-51.
- [35] Lillicrap D. Disseminated intravascular coagulation in patients with 2019-nCoV pneumonia. *J Thromb Haemost JTH*. 2020;18(4):786-87.
- [36] Giannis D, Ziogas IA, Gianni P. Coagulation disorders in coronavirus infected patients: COVID-19, SARS-CoV-1, MERS-CoV and lessons from the past. *J Clin Virol*. 2020;127:104362.
- [37] Levi M, Poll T van der. Coagulation and sepsis. *Thromb Res*. 2017;149:38-44.
- [38] Gupta N, Zhao YY, Evans CE. The stimulation of thrombosis by hypoxia. *Thromb Res*. 2019;181:77-83.

**PARTICULARS OF CONTRIBUTORS:**

1. Assistant Professor, Department of Biochemistry, Tirunelveli Medical College, Tirunelveli, Tamil Nadu, India.
2. Associate Professor, Department of Pathology, Tirunelveli Medical College, Tirunelveli, Tamil Nadu, India.
3. Professor, Department of General Surgery, Tirunelveli Medical College, Tirunelveli, Tamil Nadu, India.
4. Professor, Department of Biochemistry, Tirunelveli Medical College, Tirunelveli, Tamil Nadu, India.
5. Scientist I, Department of Multi-Disciplinary Research Unit, Tirunelveli Medical College, Tirunelveli, Tamil Nadu, India.
6. Professor, Department of General Medicine, Tirunelveli Medical College, Tirunelveli, Tamil Nadu, India.
7. Professor, Department of Pathology, Tirunelveli Medical College, Tirunelveli, Tamil Nadu, India.

**NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:**

Shantaraman Kalyanaraman,  
High Ground, Palaymkottai, Tirunelveli, Tamil Nadu, India.  
E-mail: shantaraman.kal@gmail.com

**PLAGIARISM CHECKING METHODS:** [Jain H et al.]

- Plagiarism X-checker: Aug 17, 2020
- Manual Googling: Oct 17, 2020
- iThenticate Software: Oct 29, 2020 (18%)

**ETYMOLOGY:** Author Origin**AUTHOR DECLARATION:**

- Financial or Other Competing Interests: None
- Was Ethics Committee Approval obtained for this study? Yes
- Was informed consent obtained from the subjects involved in the study? Yes
- For any images presented appropriate consent has been obtained from the subjects. NA

Date of Submission: **Aug 08, 2020**Date of Peer Review: **Sep 23, 2020**Date of Acceptance: **Oct 19, 2020**Date of Publishing: **Nov 01, 2020**